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Development of 27 novel cross-species microsatellite markers for the endangered *Hucho bleekeri* using next-generation sequencing technology

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Abstract In this study, we developed 27 polymorphic microsatellite loci in *Hucho bleekeri*, a glacial relict and freshwater resident salmonid fish in China. The number of alleles varied from 2 to 8 for each primer set. The observed and expected heterozygosity ranged from 0.250 to 0.906 and 0.508 to 0.845, respectively. The polymorphic information content ranged from 0.371 to 0.808. These microsatellite loci should be useful to study population genetics, paternity identification, speciation and adaptive evolution of this lineage.

Keywords *Hucho bleekeri* · Microsatellite · Endangered species

Hucho bleekeri, a glacial relict and freshwater resident salmonid fish, is endemic to the Yangtze River drainage in China. It became a critically endangered species mainly due to over-fishing and habitat destruction since 1960s (Hu et al. 2008). A series of measures have been taken to protect this species such as listing it into China Red Data Book to ban on fishing and establishing a national nature reserve (Wang 1998). And recently it has been performed successfully controlled breeding, which has great significance to the species preservation. To better address the

conservation status of the endangered species, a management programme based on genetics is being devised to define the efficiency of these conservation measures (Zhu et al. 2005). Despite the increasing demand for preservation and management plans, no suitable high resolution molecular markers have been available for this species. In current study, we report a set of polymorphic microsatellite loci for *H. bleekeri* that can serve as effective genetic markers for conducting further preservation and management in this endangered fish.

We used the microsatellite makers isolated from closely related species *Brachymystax lenok*, that had been constructed DNA Library by Ion PGM™ sequencer with Ion 318 chip kit v2 (Life tech LTD) in previous study (unpublished). 72 primer pairs were designed using Primer Premier 5.0 based on the flanking sequence of the repeat motifs. The polymorphisms of the loci were analyzed using a population of 24 wild individuals collected from Taibai River in Baoji City, Shaanxi Province, China. Total genomic DNA of *H. bleekeri* was extracted from tissue using the rapid salt-extraction method (Salah and Iciar 1997). The PCR amplification was carried out in a 25 µL volume that contained 1× PCR buffer, 50–100 ng genomic DNA, 0.25 µM of each primer, 200 µM dNTPs, 0.5 mM MgCl₂, and 0.25 U Taq DNA polymerase (TaKaRa). The PCR cycle comprised an initial denaturation at 94 °C for 5 min, followed by 35 cycles of at 94 °C for 30 s, at the appropriate temperature (Table 1) for 30 s and extension at 72 °C for 60 s, and then 72 °C for 10 min. The PCR products were separated on 12 % non-denaturing polyacrylamide gel and visualized by silver staining. A 20-bp DNA ladder molecular weight marker (TaKaRa) was used as the standard to determine the size of the alleles.

Among these primer sets synthesized, only 27 sets showed clearly polymorphism (Table 1 and Appendix).

Ke Wang and Shuhuan Zhang have contributed equally to this work.

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Table 1 Characterization of 27 polymorphic microsatellite loci in *H. bleekeri*

Locus	Repeat motifs	Primer sequence (5′–3′)	T _m (°C)	Size range (bp)	Na	PIC	Ho	He	P-HWE	Accession no.
BLT2	(AC)11	F: ATGACGCCACTATGAGACCG R: CAGGAAGAAGAACCAGAACCA	60	230–310	4	0.681	0.656	0.742	0.018	KM051898
BLT4	(TG)11	F: TGAACAGACACTCACACAGGC R: GTGTTTCAGCTGCTGCGTT	55	185–215	4	0.696	0.750	0.744	0.036	KM051900
BLT20	(GT)17	F: CCGTACTGCCTAGCAACACA R: GGCTGTTTTACAGAAAGGC	56	205–255	4	0.677	0.719	0.738	0.316	KM051916
BLT19	(CA)15	F: GTTCTCTCTGTCCCCTTCC R: AAACACCATGGAACCTCGACC	55	133–185	5	0.718	0.818	0.792	0.009*	KM051915
BLT6	(GT)13	F: CCCAGGTCTGGTGTCCAGTA R: GTACCGCGCTCAGCTCAT	55	105–123	4	0.696	0.708	0.760	0.024	KM051902
BLT27	(TC)11	F: GGGCAAGGTGTTATGGCTAA R: ATGAAACAGGTCCATAGCGG	58	135–153	4	0.691	0.719	0.751	0.501	KM051923
BLT28	(GT)18	F: CCCTACCAAGCACCAATACC R: TGATGTCAGGTTGCTTATTCAGA	56	155–200	5	0.746	0.563	0.794	0.003*	KM051924
BLT25	(TG)14	F: AATGAAACCAGCTCATTGCC R: CAAGTCCTTCCAAATGGTCC	56	160–200	3	0.534	0.300	0.647	0.008*	KM051921
BLT30	(TG)14	F: CCGCTCACTTTGTTGACGTA R: CCCTGTCCAACCTCTCTCAG	56	165–200	5	0.740	0.531	0.788	0.001*	KM051926
BLT16	(CA)17	F: ATCCAGTCAATAACCGCTGG R: CCTCGAGAAACTCGTTGT	55	183–205	4	0.692	0.625	0.764	0.019	KM051912
BLT31	(AG)12	F: TGGATGGGTGTTACAAGCAA R: CAGATCTTGAGACAAAGAGCCA	56	90–100	3	0.505	0.833	0.594	0.142	KM051927
Hbl1-101	(CA)13	F: TGTAATGTCACACACGCACG R: CCAGACCAGAGGGACTTCAA	55	190–240	4	0.673	0.625	0.742	0.220	KM385538
Hbl2-2	(TTA)10	F: TCTAGTGCTGCATGTCTGCC R: AGGCTCACATTGGCTCGTAT	57	140–174	5	0.741	0.625	0.789	0.199	KM385539
Hbl3-5	(TAA)11	F: TTGAAGTTGCCCTTCTGGTCC R: GGCCACACATGCAAAACAT	58	152–185	4	0.703	0.531	0.761	0.009*	KM385540
Hbl4-10	(ATC)12	F: GACAACAGCTACAGGGCACA R: GACCTGGCTCTGGGTGATAG	56	215–240	5	0.737	0.688	0.784	0.194	KM385541
Hbl5-11	(TAT)11	F: TATGTGCCCAAAATGCTGTC R: AATGGGATGTATGGGACACG	58	140–210	4	0.676	0.594	0.739	0.005*	KM385542
Hbl6-240	(AC)22	F: GGGGAATGCAGTTGAAATGT R: ATGGCATGTGTGGTGTGTTT	55	140–180	6	0.808	0.531	0.845	0.001*	KM385543
Hbl7-290	(GT)14	F: ACTCTGATCGCTACCTGGGG R: TCTGTTCGACTGCTACATCA	56	100–160	6	0.781	0.689	0.821	0.134	KM385544
Hbl8-163	(GT)15	F: GACTGGTGAGTCACAGGCAA R: GCCTAGAGTGAGACCGATGC	60	83–105	8	0.800	0.750	0.833	0.001*	KM385545
Hbl9-643	(TG)14	F: GGCACAGCATGTCCCTTTTA R: CTGCAGCATGTTCTGGATGT	55	100–125	4	0.702	0.818	0.766	0.031	KM385546
Hbl10-16	(TAA)10	F: TTCCTCTCCTCCTCCTCCAT R: GACTTCGGGGATGGCTCTAT	56	155–185	5	0.749	0.719	0.796	0.063	KM385547
Hbl11-21	(TCAT)10	F: CATTAAATCCATCCAACCATGC R: ACATCCCTGCCTTCGAAAC	56	125–200	5	0.741	0.625	0.789	0.495	KM385548
Hbl12-23	(AGTT)13	F: AATGCTTATTCACGCGAGGT R: ACACACAGCTTGGGACACAG	56	180–220	2	0.371	0.250	0.508	0.036	KM385549
Hbl13-25	(TGAG)13	F: TTTCACTGTGACCTGCTGCT R: CTCTCAACCAACACACACCG	56	88–135	5	0.748	0.688	0.796	0.074	KM385550

Table 1 continued

Locus	Repeat motifs	Primer sequence (5'–3')	T _m (°C)	Size range (bp)	Na	PIC	H _O	H _E	P-HWE	Accession no.
Hbl14-29	(TGTA) ₁₀	F: GAGGTGCACGTGTTCAAAGA R: TGGTTAAGACCAAGACCAACG	55	178–190	4	0.697	0.594	0.756	0.047	KM385551
Hbl15-31	(GTTA) ₁₀	F: GAGCTGGCTTGGTTGGTTAG R: GCACCAGTCTTCTTTTCGG	57	120–200	5	0.727	0.906	0.778	0.010	KM385552
Hbl16-128	(GT) ₁₃	F: AGCCTGTCAGCACTGATGATT R: TGGTCTGGCCTATGAACACA	52	100–120	4	0.696	0.594	0.756	0.093	KM385553

T_m annealing temperature, Size range (bp) allele range, Na observed number of alleles, PIC polymorphic information content, H_O observed heterozygosity, H_E expected heterozygosity, P-HWE probability of Hardy-Weinberg equilibrium

* $P < 0.01$, significant departures from Hardy-Weinberg equilibrium

The polymorphic parameters, including the number of alleles (Na), observed heterozygosity (H_O), expected heterozygosity (H_E), and Hardy–Weinberg equilibrium (HWE) test, for each locus were performed using Popgen32 software. And the polymorphic information content (PIC) was calculated using PIC_CALC (Cheng et al. 2013). The number of alleles per locus varied from 2 to 8. H_O and H_E were 0.250–0.906 and 0.508–0.845, respectively (Table 1). The polymorphic information content (PIC) was 0.371–0.808 (Table 1). Eight loci (BLT19, BLT28, BLT25, BLT30, Hbl3-5, Hbl5-11, Hbl6-240, Hbl8-163) deviated significantly from the Hardy–Weinberg equilibrium ($P < 0.01$; Table 1). This is the first time to develop

polymorphic microsatellite markers for *H. bleekeri* and the markers are expected to be useful for further studies of genetic diversity and paternity identification.

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Appendix: Sequences of *H. bleekeri*

